

# Conservation genetics and population history of the threatened European mink *Mustela lutreola*, with an emphasis on the west European population

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## Abstract

In species of great conservation concern, special attention must be paid to their phylogeography, in particular the origin of animals for captive breeding and reintroduction. The endangered European mink lives now in at least three well-separated populations in northeast, southeast and west Europe. Our aim is to assess the genetic structure of these populations to identify 'distinct population segments' (DPS) and advise captive breeding programmes. First, the mtDNA control region was completely sequenced in 176 minks and 10 polecats. The analysis revealed that the western population is characterized by a single mtDNA haplotype that is closely related to those in eastern regions but nevertheless, not found there to date. The northeast European animals are much more variable ( $\pi = 0.012$ ,  $h = 0.939$ ), with the southeast samples intermediate ( $\pi = 0.0012$ ,  $h = 0.469$ ). Second, 155 European mink were genotyped using six microsatellites. The latter display the same trends of genetic diversity among regions as mtDNA [gene diversity and allelic richness highest in northeast Europe ( $H_E = 0.539$ ,  $R_S = 3.76$ ), lowest in west Europe ( $H_E = 0.379$ ,  $R_S = 2.12$ )], and provide evidences that the southeast and possibly the west populations have undergone a recent bottleneck. Our results indicate that the western population derives from a few animals which recently colonized this region, possibly after a human introduction. Microsatellite data also reveal that isolation by distance occurs in the western population, causing some inbreeding because related individuals mate. As genetic data indicate that the three populations have not undergone independent evolutionary histories for long (no phylogeographical structure), they should not be considered as distinct DPS. In conclusion, the captive breeding programme should use animals from different parts of the species' present distribution area.

**Keywords:** conservation biology, microsatellites, mitochondrial DNA, *Mustela lutreola*, phylogeography, population genetics

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## Introduction

The European mink (*Mustela lutreola*) is one of the most threatened carnivores (Baillie & Groombridge 1996; [http://](http://www.redlist.org)

[www.redlist.org](http://www.redlist.org)). Its distribution started to fragment during the 19th century, and populations continue to decline dramatically (Van Bree & Saint Girons 1966; Camby 1990; Maran 1992; Tumanov 1992; de Bellefroid 1999; Sidorovich 2000; Maizeret *et al.* 2002). Presently, it lives in well-separated populations: in Estonia, Belarus and Russia [Northeast (NE) Europe; Sidorovich 2000], in Romania, namely the Danube

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delta [southeast (SE) Europe; Gotea & Kranz 1999] and finally in northern Spain (Ruiz-Olmo & Palazón 1991) and southwestern France [west (W) Europe; Van Bree & Saint Girons 1966; Camby 1990].

Understanding the population history of European mink is a key part in the conservation effort, with studies ever more urgent because of the continuing decline of the species, and captive breeding/reintroduction programmes that are already underway in various European countries (Spain, Italy, France, Germany, Czech Republic, Estonia etc.). Locally, the French restoration plan (Anonymous 1999) has proposed to release captive-bred individuals into the wild as a reinforcement measure if the other restoration measures do not appear sufficient, so the choice of the animals to be bred is of great importance. One possibility is that if the western population is genetically distinct from the eastern ones, and if the animals are locally adapted, then outbreeding depression (Lynch 1991) could result from breeding between them. On a European scale, the captive breeding programme has the stated aim to 'maintain in European zoos and other breeding facilities a population capable to maintain 90% of its heterozygosity for 100 years' (<http://www.lutreola.ee/index.html>). Captive-bred minks have already been released on Hiiumaa Island (Estonia). Thus, it is imperative that informed decisions are made regarding their management, based at least in part on genetic data.

In a preliminary study (Michaux *et al.* 2004b), we used the complete mitochondrial control region to investigate variation across a large part of the extant range of the European mink, including samples from W and NE Europe plus two individuals from Romania. We concluded that European mink probably colonized Europe from a single refugium after the last glaciation (Michaux *et al.* 2004b) because W European populations were fixed for a single haplotype. Despite low genetic differentiation between the studied populations, following the precautionary principle, we suggested that mink from the three geographically separate populations should be managed separately.

To confirm these results and be able to make more specific management advice, we conducted a new study on a larger sample, particularly for W Europe, using mitochondrial DNA markers and for the first time, nuclear microsatellites. More specifically, we tried to answer the following questions: (i) Are the three geographically separated populations genetically differentiated? (ii) Does the level of genetic diversity differ between populations? (iii) Have these populations been stable through time or have they survived recent bottleneck? (iv) Are the results using mitochondrial and microsatellites markers congruent? Specifically for the French samples, we used the microsatellite data to determine if (i) genetic substructure exists; (ii) there is evidence for isolation by distance, and (iii) genetic differentiation is associated with drainage basin or rivers.

## Methods

### Samples

A total of 176 European mink were studied, 109 from France, 15 from Spain, 34 from Romania (Danube delta), 3 from Estonia, 2 from Belarus (Vitebsk) and 13 from Russia (Tver and Pskov). The references and the geographical origin of these specimens are given in the Appendix and Fig. 1. In the mitochondrial analysis, they were compared with 10 polecats (*Mustela putorius*), two steppe polecats (*Mustela eversmannii*) and two black-footed ferrets (*Mustela nigripes*).

French, Spanish and Romanian samples were collected either from animals caught, marked and released, or found dead. These tissues are conserved in the tissue collection of J. R. Michaux. The other samples were described previously in Davison *et al.* (2000).

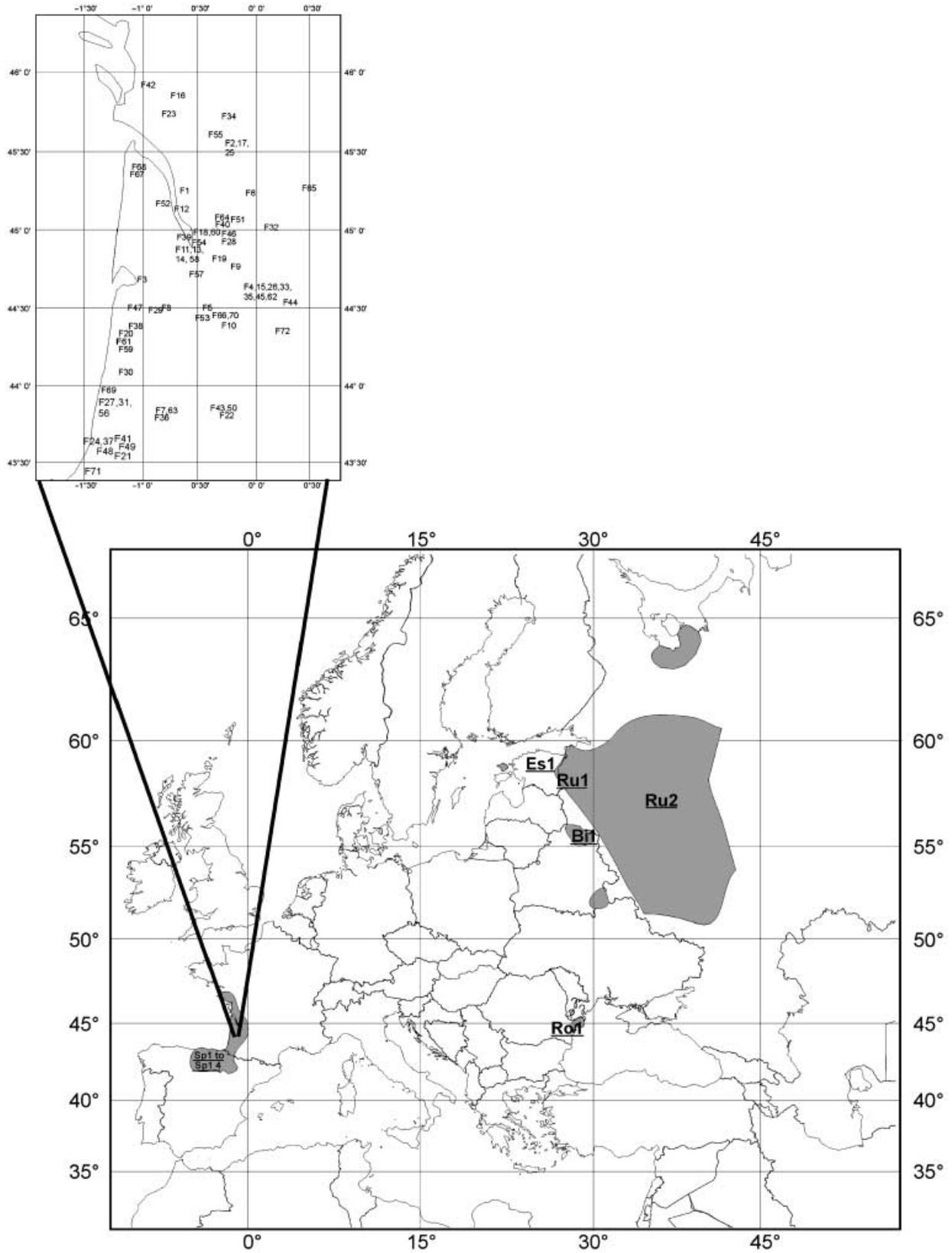
### DNA methods

DNA was extracted from ethanol-preserved tissue using the phenol-chloroform method as described by Sambrook *et al.* (1989).

**Mitochondrial DNA.** The complete mitochondrial control region was amplified using specific primers L0ML (5'-TAT TCTAACTAACTATTCCTG-3') and EML (5'-CTA TAGATGTRT TTATAACCC-3'). Amplification reactions were carried out in 2 × 50 µL volumes including 25 µL of each 2 µM primer, 20 µL of 1 mM dNTP, 10 µL of 10× Promega reaction buffer B, 2.5 mM MgCl<sub>2</sub>, 10 µL of purified water and 0.2 µL of 5 U/µL Promega *Taq* DNA polymerase. Approximately 200 ng of DNA (10 µL) was used per PCR amplification. PCR was performed using an MJ Research PTC100 thermal cycler, employing 33 cycles (20 s at 94 °C, 30 s at 50 °C and 1 min 30 s at 68 °C) with a final extension cycle of 10 min at 68 °C. PCR products were then purified using the Ultra-free DA Amicon kit (Millipore) and directly sequenced. Both strands were sequenced using a BigDye terminator sequencing kit (Applied Biosystems) on an ABI 310 (Applied Biosystems) automated sequencer.

The newly determined sequences from 171 European minks were compared with five European mink (AF207720, AF207721, AF207723, AF207724 and AF207725) and 10 polecat (AF068570, AF207717, AF207718 and AF207726; AJ548803 to AJ548808) partial control region sequences available in GenBank (Appendix). Sequences were aligned using the ED editor (MUST package; Philippe 1993).

**Microsatellites.** Pairs of microsatellite primers were selected from previous studies on mustelids (*M. vison*: Fleming *et al.* 1999; *M. erminea*, *Martes pennanti*, *Martes americana*, *Lutra canadensis*: Davis & Strobeck 1998; Fleming *et al.* 1999). In a first step, 20 pairs of primers were tested on 10 European



**Fig. 1** Geographic distribution of the *Mustela lutreola* samples (see Appendix for the sample symbols). The shaded zones correspond to the distribution area of the species (according to T. Maran, personal communication).

mink originating from different parts of the distribution area. Positive results were obtained with 18 of them (Mvis002, Mvis020, Mvis022, Mvis072, Mvis075, Mvis099, Mvis92534, Mer005, Mer009, Mer022, Mer 41, Mer095, Ma1, Ma10, Ma19, GG7, GG-14, TT4), so these were then tested on a greater sample (five individuals from each population). Six loci (Mvis020, Mvis072, Mvis075, Mer009, Mer022, Mer 41) were polymorphic and were scored on a total sample of 155 individuals.

Amplification reactions were carried out in 15  $\mu$ L volumes including  $\sim$ 10 ng of DNA template, 24  $\mu$ M of each dNTP, 1 mM MgCl<sub>2</sub>, 3.5 pmoles of the [ $\gamma$ -<sup>33</sup>P]-dATP labelled forward primer, 15 pmoles of the unlabelled reverse primer, 0.75 unit of *Taq* DNA polymerase (Goldstar, Eurogentec), 75 mM Tris-HCl pH 9, 20 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.01% (w/v) Tween 20. PCR was performed using a PTC 100 thermal cycler (MJ Research); 94 °C for 5 min, followed by 94 °C for 1 min, 30 s at annealing temperature (varying according to the microsatellite), 72 °C for 1 min for 30 cycles, final extension at 72 °C for 5 min. Amplification products (15  $\mu$ L) were mixed with 6.7  $\mu$ L formamide loading dye and then electrophoresed in 6% acryl-bisacrylamide and 8 M urea sequencing gels for 3–5 h at about 1800 V. Sequencing reactions of pUC19 vector (Appligene) were also loaded adjacent to the samples, to serve as a size standard. Once dried, gels were exposed to X-ray films for 24–48 h.

### Data analyses

**Mitochondrial DNA.** The aligned sequences were analysed by distance (neighbour joining, NJ; Saitou & Nei 1987), maximum-parsimony (MP; Fitch 1971) and maximum-likelihood (ML) methods using PAUP 4.0b8 (Swofford 1998). The general time reversible (GTR) model and Kimura 2-parameter (K2P) estimator were used for the distance and ML analyses (chosen with MODELTEST 3.0, Posada & Crandall 1998). These analyses were developed assuming a gamma distribution for substitution rates across sites, where the parameter alpha (Yang 1996) and the proportion of invariant sites (*I*) were estimated with the ML method using PAUP. MP analyses were conducted according to a heuristic search and TBR branch swapping option. The robustness of inferences was assessed by bootstrap re-sampling (BP) 1000 random repetitions for MP and distance analyses, and 100 for ML.

A Bayesian approach to phylogeny reconstruction (Yang & Rannala 1997; Huelsenbeck *et al.* 2001) was also used, implemented in MRBAYES 2.01 (Huelsenbeck *et al.* 2001). Metropolis-coupled Markov chain Monte Carlo sampling was performed with four chains that were ran for 500 000 generations, using default model parameters as starting values. Bayesian posterior probabilities were picked from the 50% majority rules consensus of trees sampled every 20 generations, after removing trees obtained before chains

reached apparent stationarity ('burn in' determined by empirical checking of likelihood values).

A network was constructed using the program tcs (Clement *et al.* 2000) as this method is very useful when sequences are closely related.

Nucleotide ( $\pi$ ) and haplotype (*h*) diversities, were estimated using the DNASP program (Rozas & Rozas 1997). Calculations were performed on the main data matrix, including 176 animals. The 'mismatch distribution' of substitutional differences between pairs of haplotypes was calculated within the NE and SE geographical groups and compared with a fit to the Poisson model using DNASP (Rozas & Rozas 1997).

Haplotype diversity was partitioned among populations by computing pairwise  $G_{ST}$  and  $N_{ST}$  (Pons & Petit 1996) between three geographical separate groups (W Europe, SE Europe, NE Europe) using the software SPAGED1 (Hardy & Vekemans 2002). Contrary to  $G_{ST}$ ,  $N_{ST}$  accounts for the phylogenetic distances between haplotypes (estimated by the minimum number of mutational events between haplotype sequences) and is expected to be larger than  $G_{ST}$  when a phylogeographical pattern occurs (i.e. when related haplotypes co-occur more often within population than random expectation). To test for a phylogeographical pattern, 1000 random permutations of haplotype identities were made, keeping the haplotype frequencies and the matrix of pairwise haplotype distances intact, and the distribution of  $N_{ST}$  values obtained after permutation was compared with the observed value (Burban *et al.* 1999). Finally, the population genetic structure was determined by analysing the molecular variance (AMOVA available in ARLEQUIN 2.000; Schneider *et al.* 2000). This method estimates the proportion of genetic variation at different hierarchical levels: among groups (corresponding to the three main mink groups: W, SE and NE Europe), among populations within each group (France and Spain for the W group; Russia, Belarus and Estonia for the NE group) and within each population. The program ARLEQUIN was also used to calculate  $\Phi_{ST}$  statistics that measure population subdivision analogous to Wright's (1992) *F*-statistics.

**Microsatellites.** Pairwise  $F_{ST}$  (Weir & Cockerham 1984) values between the three geographical groups, were computed using FSTAT version 2.9.3 (Goudet 1995). We compared the levels of genetic diversity among these groups by their gene diversity ( $H_E$ ) and allelic richness ( $R_S$ ) using FSTAT.  $R_S$  is the mean number of alleles expected within a sample of defined size (in practice the size of the smallest group) and is therefore appropriate to compare allelic richness when sample size varies.

A phylogenetic tree was also constructed on the basis of the microsatellite markers polymorphism using the TREEMAKER 2.0 program (Piry, personal communication). For this, the BIONJ algorithm (Gascuel 1997) and the distance

model of Cavalli-Sforza & Edwards (1967) were used. The robustness of inferences was assessed by bootstrap resampling (1000 random repetitions).

To discover whether there is a signature of recent bottleneck events, we used the software BOTTLENECK (Cornuet & Luikart 1996) which compares the gene diversity observed ( $H_E$ ) with the one expected from the number of alleles per locus ( $A_0$ ) when population size remains constant and for a given mutation model. After a bottleneck, one expects that  $A_0$  drops more than  $H_E$ , so that the observed  $H_E$  should be higher than that expected on the basis of  $A_0$ . As the mutation model of microsatellites is thought to be intermediate between a stepwise-mutation model (SMM) and an infinite allele mutation model (IAM), we tested bottleneck events assuming each of these models. We used the Wilcoxon signed rank test to test significance as suggested by Cornuet & Luikart (1996).

The inbreeding coefficient,  $F_{IS}$ , describing how heterozygote frequencies deviate from expectations under panmixia within defined subpopulations, was estimated for each geographical group. In NE Europe, sample sites were considered as different subpopulations because they are quite distant, whereas a single 'subpopulation' was assumed in W Europe and in SE Europe. To test  $F_{IS}$ , genes were randomized within subpopulations, and loci were jackknifed to estimate standard errors. Computations were carried out using the software SPAGED1 (Hardy & Vekemans 2002).

To characterize the genetic structure within the W European population (actually the French population as only three Spanish samples could be genotyped), we used SPAGED1 to compute pairwise kinship coefficients between individuals ( $F_{ij}$ ) using Nason's multilocus estimator defined in Loiselle *et al.* (1995).  $F_{ij}$  values were regressed on  $\ln(d_{ij})$ , where  $d_{ij}$  is the geographical distance between the sampled locations of individuals  $i$  and  $j$ , and the regression slope ( $b$ ) was used to quantify the extent of isolation by distance. To test for a nonrandom spatial structure, we applied a Mantel test between the matrices of  $F_{ij}$  and  $\ln(d_{ij})$  values using 10 000 randomizations.  $F_{ij}$  values were also averaged over a set of mutually exclusive distance classes, giving  $F(d)$  values. To determine whether gene flow occurs preferentially within drainage basins, two  $F(d)$  curves were computed: one for  $i$ - $j$  pairs sampled within a same drainage basin and one for  $i$ - $j$  pairs sampled in different basins.

## Results

### Mitochondrial DNA

**Sequences.** The complete sequence of the mitochondrial control region was obtained for 171 European minks and associated to five gene bank sequences (see above). These sequences coming from 50 different localities were added to 10 European polecats from five regions, two steppe

polecats and two black-footed ferrets (*Mustela nigripes*), used as an outgroup (Michaux *et al.* 2004) (Table A1). The different haplotypes observed in this data matrix have been deposited in the EMBL GenBank under accession nos AJ548474 to AJ548477 and AJ548803 to AJ548822.

**Phylogenetic analyses.** The analyses were performed, using the complete mitochondrial control region, with two *Mustela nigripes*, two *Mustela eversmannii*, 10 *Mustela putorius* and 176 *Mustela lutreola*. All the French and Spanish animals share the same mtDNA haplotype with the exception of an hypervariable  $C_nT_n$  array (either 7 or 8 thymine residues) and a variable (from 11 to at least 23) number of copies of an 11 bp minisatellite in the hypervariable region R. It was impossible to determine the exact number of minisatellite copies, when  $n > 23$ , due to sequencing difficulties, so this region was not considered in any further phylogenetic analysis.

After removal of the  $C_nT_n$  array and the minisatellite repeats, the final data matrix involved 25 different haplotypes and 731 sites, of which 64 were variable and 43 phylogenetically informative. The mean estimated transition to transversion ratio was 3.0 and the nucleotide frequencies were C 26.3%, T 29.4%, A 27.9%, and G 16.3%.

The neighbour-joining tree (Fig. 2) shows two major clades: the first one corresponding to the polecat, *M. putorius*, and the second subdividing into two monophyletic groups of *M. lutreola* and *M. eversmannii*. The Bayesian, ML and MP (one most parsimonious tree, L = 86 steps; CI = 0.68; RI = 0.81) analyses yielded phylogenies of identical structure. Bootstrap values and Bayesian probabilities (BaP) resulting from these analyses are indicated in Fig. 2. The group corresponding to *M. putorius* is well supported (BP values for NJ: 71%, MP: 78, ML: 73% and BaP: 0.95). The second group of *M. lutreola* and *M. eversmannii* is not supported (BP values for NJ: 38%, ML: 52% and BaP: 0.42). No genetic structure associated to geography is observed within *M. lutreola*.

The network analysis (Fig. 3) shows a similar result, i.e. a clear separation between the three species *M. eversmannii*, *M. putorius* and *M. lutreola* and within this last species, a total absence of genetic structure associated to geography.

**Genetic structure.** Nucleotide ( $\pi$ ) and haplotype ( $h$ ) diversities were calculated for each population of *M. lutreola* (Table 1). Animals from NE Europe (Russia, Estonia, Belarus) had a high nucleotide and haplotype diversity, as compared to the W European samples (France Spain), which shared a single mitochondrial type. The 34 SE European (Romania) animals are characterized by four different haplotypes, so the nucleotide and haplotype diversities are intermediate.

As already observed in Michaux *et al.* (2004), the mismatch distribution of pairwise differences showed a signature of population growth (bell-shape) for the NE European

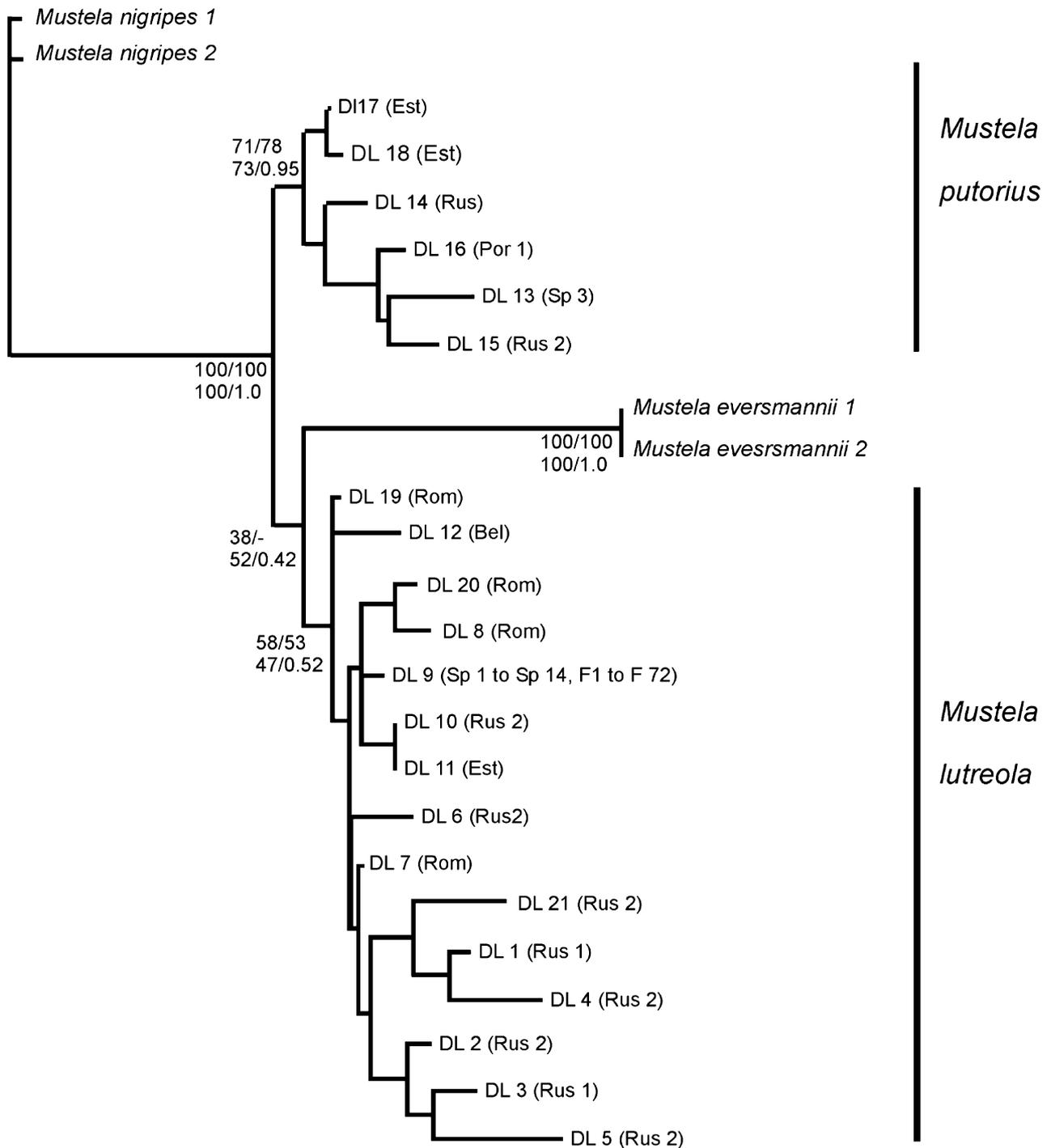
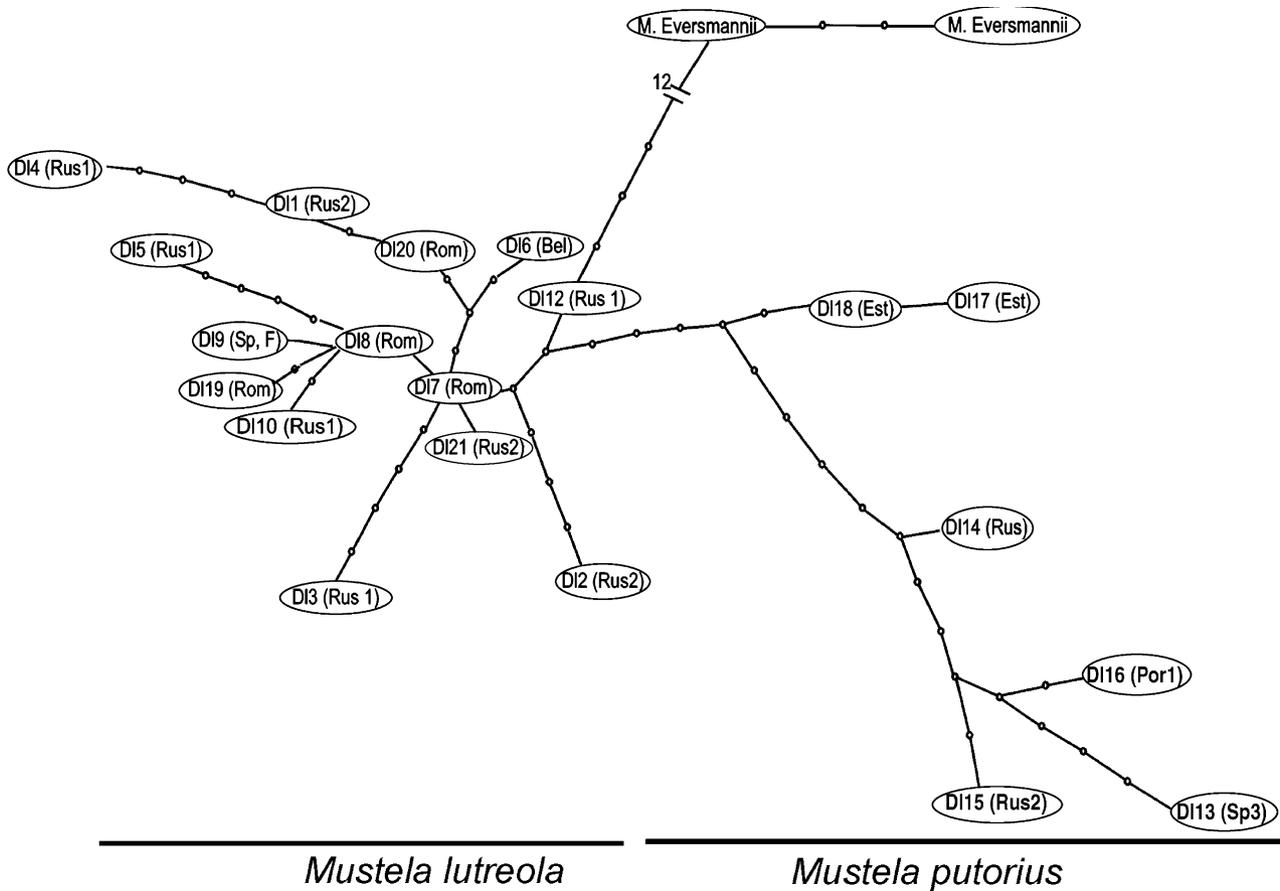


Fig. 2 Consensus neighbour-joining tree derived from the analysis of the complete mitochondrial control region sequences for 121 European mink, polecats and black-footed ferrets (used as outgroup). Each haplotype is identified by the letters DL + a specific number. The locality codes (see Appendix) are also given for each haplotype. For each well supported node, the different robustness are indicated as followed: neighbour joining/maximum parsimony; maximum likelihood/Bayesian probability.

group (Fig. 4a). On the contrary, the SE European population was characterized by a signature of constant size population (Fig. 4b). Pairwise  $G_{ST}$  values between geographical groups were all significant ( $P < 0.001$ ) and are of

0.89 between SE Europe and W Europe; 0.54 between W Europe NE Europe and 0.42 between this last region and SE Europe.  $N_{ST}$  were lower than  $G_{ST}$  in all pairwise comparisons (respectively 0.84, 0.39 and 0.22) and haplotype



**Fig. 3** TCS network constructed using mitochondrial control region sequences. The identity of the haplotypes (see Fig. 2) and their geographical origin (see Appendix) are indicated. The open squares between haplotypes correspond to intermediate haplotypes as calculated by the tcs program. The number of intermediate haplotypes between *M. lutreola* and *M. eversmannii* being high (12), all of them were not indicated and were replaced by the number 12.

**Table 1** Genetic variability observed within the three populations of *Mustela lutreola* using the mitochondrial control region

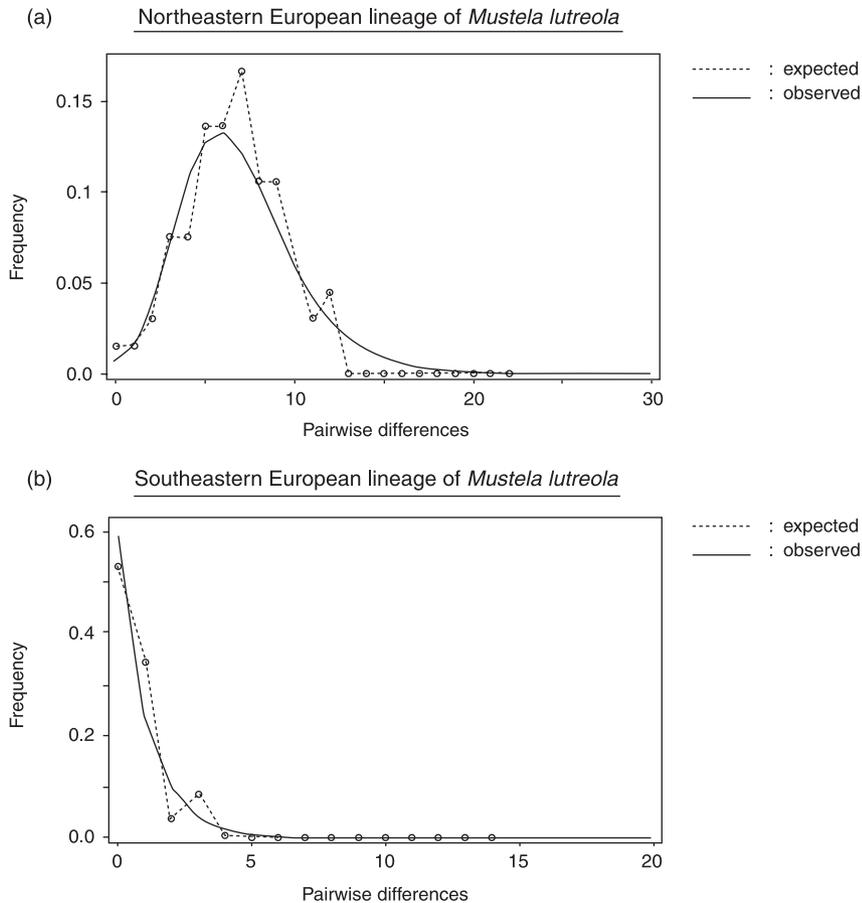
|                  | Sample size | Number of haplotypes | Genetic divergence within each group (% K2P distance) | Nucleotide diversity ( $\pi$ ) | Standard deviation | Haplotype diversity ( $h$ ) | Standard deviation |
|------------------|-------------|----------------------|---|--------------------------------|--------------------|-----------------------------|--------------------|
| Northeast Europe | 18          | 10                   | 1.5   | 0.012                          | 0.0014             | 0.939                       | 0.058              |
| Southeast Europe | 34          | 4                    | 0.3   | 0.0012                         | 0.0003             | 0.469                       | 0.088              |
| West Europe      | 124         | 1                    | 0   | 0                              | /                  | 0                           | /                  |

identity permutation tests were always nonsignificant. Hence, there is no evidence of a phylogeographical structure. All  $\Phi_{ST}$  values were also significant ( $P < 0.001$ ) (respectively 0.91, 0.71 and 0.26) and on the contrary to the two other indices, it indicates (high value of  $\Phi_{ST}$ ) a separation between W Europe and the two oriental populations (SE and NE Europe). In contrast,  $\Phi_{ST}$  values are low between these two last populations indicating a closer relationship. The AMOVA shows that a moderate percentage (25%) of the total mtDNA variation is distributed among

the three genetic groups whereas a higher value of this variation (48%) is observed among populations within the main lineages. This strongly indicates a weak phylogeographical structure for this species.

*Microsatellites*

As it was impossible to amplify several microsatellite markers for some specimens, only 155 European minks were used for the microsatellite analyses. Of the 18 microsatellite



**Fig. 4** Mismatch distribution for mtDNA types from the Russian–Estonian–Belarus and Romanian genetic lineages of *Mustela lutreola*. The expected frequency is based on population growth–decline (a) or constant (b) models (For the groups, respectively:  $\theta$  initial = 0.82 and 0.40,  $\theta$  final = 1000,  $\tau$  = 5.73 and 0.29), determined using the DNASP 3.5 program (Rozas & Rozas 1997).

loci tested, only six were polymorphic, containing up to five alleles (Table 2). One locus displayed an extreme heterozygote deficit in the NE Europe sample, likely due to a null allele, and was therefore removed from our analyses. Of the total of 20 alleles at all five loci, 19 were found in the NE Europe sample ( $N = 19$  individuals), 15 in SE Europe ( $N = 25$ ), and 13 in W Europe ( $N = 112$ ). The same trend was found for gene diversity ( $H_E$ ) and allelic richness ( $R_S$ ; Table 3). Permutation tests performed by the FSTAT software confirm that W Europe has less genetic diversity than NE Europe on the basis of  $H_E$  ( $P = 0.021$ ) and  $R_S$  ( $P = 0.003$ ), whereas SE Europe lies in between ( $H_E$  and  $R_S$  values not statistically different from the ones found in W or NE Europe). The mean values of  $H_E$  for the European mink are significantly lower than those observed on other aquatic mustelids such as the otter (*Lutra lutra*) (average of 0.74; Randi *et al.* 2003) but is of the same order of what is observed in the wolverine (*Gulo gulo*) (from 0.269 to 0.376; Walker *et al.* 2001).

The Wilcoxon signed rank tests performed by the BOTTLENECK software, which test for recent bottleneck events, were nonsignificant in NE Europe whatever the mutation model assumed, but significant in W and SE Europe using the IAM (Table 2). Under the SMM, the test was near

**Table 2** Allele frequencies at six microsatellite loci within each region sampled

| Locus   | Allele (PCR product size) | West Europe ( $N = 112$ ) | Southeast Europe ( $N = 25$ ) | Northeast Europe ( $N = 18$ ) |
|---------|---------------------------|---------------------------|-------------------------------|-------------------------------|
| Mvis075 | 142                       | 0.396                     | 0.920                         | 0.763                         |
|         | 144                       | 0.604                     | 0.080                         | 0.237                         |
| Mer009  | 196                       | 0.116                     | 0.120                         | 0.206                         |
|         | 202                       | 0.000                     | 0.000                         | 0.265                         |
|         | 204                       | 0.000                     | 0.120                         | 0.000                         |
|         | 206                       | 0.884                     | 0.380                         | 0.412                         |
| Mvis72  | 208                       | 0.000                     | 0.380                         | 0.118                         |
|         | 268                       | 0.000                     | 0.000                         | 0.026                         |
|         | 272                       | 0.677                     | 0.860                         | 0.605                         |
|         | 274                       | 0.000                     | 0.140                         | 0.263                         |
| Mer022  | 278                       | 0.323                     | 0.000                         | 0.105                         |
|         | 243                       | 0.000                     | 0.000                         | 0.194                         |
|         | 251                       | 0.005                     | 0.413                         | 0.389                         |
|         | 253                       | 0.269                     | 0.239                         | 0.167                         |
| Mer41   | 255                       | 0.000                     | 0.022                         | 0.222                         |
|         | 257                       | 0.726                     | 0.326                         | 0.028                         |
|         | 159                       | 0.766                     | 0.587                         | 0.861                         |
|         | 161                       | 0.220                     | 0.000                         | 0.056                         |
|         | 163                       | 0.009                     | 0.391                         | 0.056                         |
|         | 165                       | 0.005                     | 0.022                         | 0.028                         |

**Table 3** Gene diversity ( $H_E$ ) and allelic richness ( $R_S$  for a sample of 17 individuals) at five microsatellite loci within each geographical group, and  $P$  values of Wilcoxon tests of gene diversity excess (indicative of past bottleneck events) assuming an infinite allele mutation model (IAM) or a stepwise-mutation model (SMM)

|          | West Europe | Southeast Europe | Northeast Europe |
|----------|-------------|------------------|------------------|
| $H_E$    | 0.379       | 0.458            | 0.539            |
| $R_S$    | 2.12        | 2.89             | 3.76             |
| Test IAM | 0.031       | 0.031            | 0.500            |
| Test SMM | 0.594       | 0.062            | 0.890            |

significant in SE Europe. Hence, there is no conclusive evidence of a past bottleneck event but hints that such an event occurred in SE and possibly W Europe.

Pairwise  $F_{ST}$  values between geographical groups were all significant ( $P < 0.001$ ):  $F_{ST} = 0.10$  between SE and NE Europe,  $F_{ST} = 0.29$  between W. Europe and SE Europe, and  $F_{ST} = 0.26$  between W Europe and NE Europe. Hence, the population from W Europe is the most differentiated. The phylogenetic tree (data not shown) constructed on the basis of the microsatellite polymorphism gave a similar result as the western European group (France and Spain) appear separated to the other eastern populations (BP = 75%).

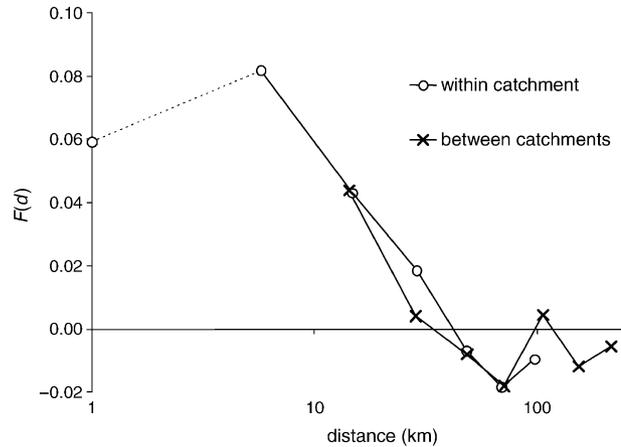
In W Europe, a clear pattern of isolation by distance was observed whereby the kinship coefficients between individuals decrease approximately linearly with the logarithm of the distance up to c. 50 km and then stabilize (Fig. 5). The genetic structure is statistically significant (Mantel test:  $P = 0.004$ ) and the regression slope of  $F_{ij}$  on  $\ln(d_{ij})$  for distances inferior to 50 km is  $b = -0.0496$ . There is no difference between the  $F(d)$  curves within and between basins (Fig. 5), indicating that gene flow is not affected by hydrography.

Within geographical groups, statistically significant heterozygote deficit was found at two loci (Mer022:  $F_{IS} = 0.192$ ,  $P = 0.006$ ; Mer41:  $F_{IS} = 0.249$ ,  $P = 0.012$ ) but not at the three other ones (Mvis075:  $F_{IS} = 0.020$ ; Mer009:  $F_{IS} = 0.071$ ; Mvis72:  $F_{IS} = 0.062$ ). Multilocus  $F_{IS}$  values reach 0.084 (SE = 0.062;  $P = 0.073$ ) in W Europe, 0.085 (SE = 0.0063;  $P = 0.28$ ) in SE Europe, and 0.182 (SE = 0.090;  $P = 0.033$ ) in NE Europe. It is worth noting that the  $F_{IS}$  in W Europe is close to the mean kinship coefficient between nearby individuals ( $F_{ij} = 0.08$  for distances < 10 km), as expected if mating occurs locally.

## Discussion

### The phylogeography of European mink

As stated in our previous study (Michaux *et al.* 2004), *Mustela lutreola* shows low mitochondrial DNA sequence



**Fig. 5** Relationship between the mean kinship coefficients between individuals of the W European population and the logarithm of the distance separating them. The two curves distinguish pairs of individuals sampled either in the same or in different basins. The symbol on the vertical axis indicates the value found for individuals sampled at the same location.

variation and little geographical partitioning of haplotypes (Table 1, Figs 2 and 3). This is consistent with other carnivores, especially mustelids like wolverine (*Gulo gulo*) (Walker *et al.* 2001), European otter (*Lutra lutra*) (Cassens *et al.* 2000; Morales 2002), polecat (*Mustela putorius*) and pine marten (*Martes martes*) (Davison *et al.* 2001), fisher (*Martes pennanti*) (Drew *et al.* 2003) as well as wolf (*Canis lupus*) (Vila *et al.* 1999). In contrast, other carnivores such as American marten (*Martes americana*) (Carr & Hick 1997), ermine (*Mustela erminea*) (Fleming & Cook 2002) and European (Taberlet *et al.* 1998) and North American brown bear (Waits *et al.* 1998) (*Ursus arctos*) have structured mtDNA phylogenies that correlate with geography. These patterns are probably largely shaped by species differences in the degree of range fragmentation during the last ice age, the level of dispersal following the withdrawal of the glaciers, and the extent of gender-bias in dispersal (Drew *et al.* 2003).

As proposed in other mustelids like the pine marten (*Martes martes*) (Davison *et al.* 2001), a general lack of ancient lineages and a mismatch distribution for the NE group that is consistent with an expanding population, suggests that the present-day *M. lutreola* colonized Europe from a single refugium following recent glaciation. This is also consistent with the lack of any phylogeographical structure according to the test of  $N_{ST}$  and AMOVA. As hypothesized by Avise (2000) and Hewitt (1996), it is expected that populations living in the southern refugial regions were less affected by climatic changes and should have more genetic variation. In contrast to other mammals, where the southern-most refugial populations have the greatest diversity (Merilae *et al.* 1997; Michaux *et al.* 2003, 2004; Hewitt 2004), European mink mtDNA and microsatellite diversities

are highest in NE European populations (Russia, Belarus, Estonia). Therefore, the present-day populations of western and southeastern Europe cannot have been the refugial populations. However, as the majority of the southern-central European populations of the mink have become extinct (Youngman 1982; de Bellefroid & Rosoux 1998; de Bellefroid 1999) and as the fossil record for this species is sparse (Davison *et al.* 2000), particularly in southern Europe, it is impossible to conclude definitively that Mediterranean regions were never a refuge for the European mink during the Pleistocene glaciations.

Nevertheless, the current data (Figs 2 and 3) confirm previous studies on mitochondrial DNA (Davison *et al.* 2000; Michaux *et al.* 2004) and indicate that European mink and steppe polecat are closely related. This means that the European mink may have colonized Europe and western Russia relatively recently from an eastern refuge, as has been inferred in other mammal species [root vole (*Microtus oeconomus*), Brunhoff *et al.* 2003].

#### *Demographic history of the European mink populations*

The almost complete lack of variation observed in French and Spanish animals strongly indicates that very few individuals established the present-day W European population, possibly following a human introduction. This hypothesis is corroborated by the absence of mink records in France until the first half of the 19th century and in Spain until 1950 (de Bellefroid 1999). As the microsatellite allelic richness of the whole W European population is extremely low, the hypothesis of a leptokurtic dispersal of some long-distance migrants establishing populations in advance of a colonization wave during the Holocene (Ibrahim *et al.* 1996) becomes a less conceivable explanation for the low diversity. The high  $F_{ST}$  between this population and the eastern ones probably results from the strong genetic drift undergone by this population, a hypothesis supported by the test of bottleneck event.

The SE European population is probably a relict of a more important population which was previously widespread all over central Europe and Ukraine. This would explain the low  $F_{ST}$  value observed between the NE and SE European groups as compared to  $F_{ST}$  between these populations and the W European one. Moreover, the isolation of the Romanian minks has probably been associated with a genetic bottleneck as indicated by (i) the low microsatellite allelic richness, (ii) the excess of microsatellite  $H_E$  as compared to the number of alleles (Table 2), and (iii) the few mtDNA haplotypes and low nucleotide diversity as compared to the Russian animals. After the bottleneck, the Romanian population stayed relatively constant as indicated by the mismatch distribution analysis (Fig. 4).

The animals from NE Europe are characterized by the highest level of genetic diversity (high number of mito-

chondrial DNA haplotypes and high microsatellite allelic richness), so have not undergone a recent bottleneck. On the contrary, the mismatch distribution analysis suggests a recent population expansion after the last glaciation, which was followed by a historical decline and fragmentation of populations. This would explain why even this NE European population is characterized by a lower level of genetic variability as compared to other aquatic Mustelidae such as the European otter *Lutra lutra*. Indeed, this last species have probably less suffered to historical extinction or fragmentation of populations (Randi *et al.* 2003).

#### *Intrapopulation structure of the western European group*

In western Europe, the genetic structure of European mink shows isolation-by-distance pattern up to *c.* 50 km. Beyond, the relatedness between individuals does not decrease any more with distance. It is congruent with field observations made in Russia, Belarus and Spain which indicate that European mink are very sedentary, rarely leaving the rivers where they were born (Maran & Ceña, personal communication). However, in SW France, radio-tracking experiments showed that two males displayed a nomadic behaviour, leaving their home range, crossing the limits of 'their' catchment and moving on a distance of more than 40 km (Fournier, unpublished). This unusual behaviour is probably the consequence of a very low population density (Camby 1990) involving the absence of any female and forcing the males to look for mates along other hydrographic systems. Therefore, we think this behaviour is not a peculiar adaptation of the French mink population but is more the consequence of an individual mating tactic adopted when demographic densities are very low.

Moreover, our study evidenced that relatedness between French individuals was not affected by the pattern of basins, indicating that French European mink does not preferentially disperse along streams and rivers, at least for reproductive purposes. This is in contrast to what it is observed in Russia, Belarus and Spain (when densities are high). Again, this would be interpreted as the result of low mink densities in France, forcing them to disperse a lot for reproduction, not only along the rivers, but also between drainage basins.

The deficit of heterozygotes observed in W Europe is consistent with the isolation-by-distance pattern as the  $F_{IS}$ , which is expected to equal the kinship coefficient between mates, is similar to the kinship coefficient between nearby individuals. Heterozygote deficit can thus be explained by limited dispersal capabilities causing related individuals to mate. Nevertheless, it cannot be excluded that null alleles also contribute to an apparent heterozygote deficit in some loci. If we assume that the pattern of isolation by distance is at drift-dispersal equilibrium for distances less

than 50 km, the neighbourhood size ( $N_b$ ) can be estimated from  $b$ , the regression slope of  $F_{ij}$  on  $\ln(d_{ij})$ :  $N_b = [F(1) - 1]/b = 18$ , where  $F(1)$  is the mean kinship coefficient between neighbouring individuals (Vekemans & Hardy 2004).  $N_b$  estimates the quantity  $4\pi D\sigma^2$ , where  $D$  is the effective population density and  $\sigma^2$  is half the mean squared parent-offspring dispersal distances. Hence, intergeneration dispersal distances could be estimated by assessing population density, but this is currently an unknown parameter. Nevertheless,  $N_b$  also provides an order of magnitude of the number of potential mates (i.e. within reach) per individual.

#### *Implications for the conservation of European mink*

The main conservation issue is whether mink populations with very restricted genetic diversity could survive for a long period of time. Several recent studies on other wild mammals [moose (*Alces alces*): Ellegren *et al.* 1993, 1996; European beaver (*Castor fibre*): Mikko & Anderson 1995; San Nicolas Island fox (*Urocyon littoralis dickeyi*): Aguilar *et al.* 2004] have demonstrated that a conservation programme, or the survival of a species, can be successful despite low levels of genetic variation in the founder population. While the short-term chance of survival may mainly depend on environmental pressures (diseases, destruction of habitats), long-term survival may be more dependent upon genetic variability especially that found at disease resistance loci (Aguilar *et al.* 2004). This was confirmed recently by Spielman *et al.* (2004) which demonstrated that threatened species presenting a lower genetic diversity have higher extinction risks than species characterized by a high genetic variability. Therefore, as the European mink is presently suffering severe environmental pressures (destruction or pollution of aquatic habitats, etc.) and introduced Aleutian disease (Fournier-Chambrillon *et al.* 2004), the best chance to save this species in the long term will be to maximize genetic variability, particularly in the captive breeding/reintroduction programmes that are presently underway as well as to continue developing habitat conservation and restoration measures.

Awise (2000) defined the concept of a management unit as 'any population that exchanges so few migrants with others as to be genetically distinct from them normally will be demographically independent at the present time'. In this way, he noted that 'even shallow matrilineal subdivisions can be relevant to conservation efforts'.

However, the value of this concept as well as others like the ESU (evolutionary significant unit, Ryder 1986) was strongly debated these last years and are often difficult to define (Fraser & Bernatchez 2001) and to apply in practice (Moritz 1994). Moreover, other authors (Crandall *et al.* 2000) disagree about the new interpretation of these concepts which are often based exclusively on molecular data not taking ecological information into account. However,

this is also extremely important to recognize specific adaptation to local environment and to avoid problems of outbreeding depression in the case of translocation or reintroduction programmes (Drew *et al.* 2003).

For this reason, we preferred to use the concept of 'distinct population segments' (DPS) proposed by the US Fish and Wildlife Service and National Marine Fisheries Services (1996) which appears more relevant. Indeed, to be considered as a DPS, a population must be (i) discrete and (ii) biologically and ecologically significant. Discrete populations are geographically isolated from other ones by physical, physiological, ecological or behavioural factors. Biological and ecological significance is determined by a variety of potential factors including the fact that a population occurs in a unique or unusual ecological setting, its loss would result in a significant gap in the range of the species, or it differs markedly from other populations in its genetic characteristics (US Fish and Wildlife Service & National Marine Fisheries Services, 1996).

As they are geographically well isolated, our mink populations are discrete. Concerning the biological significance, on the genetic point of view, certainly, specific mitochondrial haplotypes are found in the three main European mink populations, with a single haplotypes characterizing the French and Spanish animals. This involves a high  $\Phi_{ST}$  value between western and eastern groups. Moreover, the microsatellite analysis showed high differentiation between the W European population and the NE, also indicating weak gene flow between them. This would tend to define them as different genetically distinct groups. However, the same microsatellite alleles are found within all three main genetic groups (W, SE and NE Europe), the AMOVA showed that a moderate percentage (25%) of the total mtDNA variation was distributed among them, the test of  $N_{ST}$  indicates a lack of any phylogeographical structure between them and the SW European haplotype is only different from several eastern ones by a single nucleotide out of 731. This indicates that these mink groups are closely related and that their particular genetic patterns are probably the result of a recent fragmentation of a more widespread population. Thus, the genetic differentiation between western and eastern populations is rather explained by a founder effect.

From an ecological point of view, it seems impossible to find any particular ecological adaptation separating the three European populations from each other. Indeed, in Spain, the mink can live either in (sub)Mediterranean habitats or in rainy mountain forests with strong climatic conditions, similar to those found in Russia (heavy rainfall and long frost periods). In France, it still lives or was present in temperate humid habitats (rivers or marshes), just like in the Romanian Danube delta.

As far as the dispersal pattern is concerned, our results indicate that the differences observed between or within

populations are more related to differences in the population densities rather than to specific adaptations associated to particular environmental conditions.

Therefore, as the three European mink populations (SW, SE and NE Europe) are discrete but do not seem to be biologically significantly isolated, we think that it is difficult to consider them as specific 'distinct population segments'. They rather correspond to a single population which was recently fragmented.

In conclusion, as already proposed for other threatened species such as the fisher (*Martes pennanti*) (Drew *et al.* 2003), the Mariana crow (*Corvus kubaryi*) (Tarr & Fleischer 1999) or the Eld's deer (*Cervus eldi*) (Balakrishnan *et al.* 2003), an interesting way to preserve the European mink in the long term should be to increase the genetic variability of impoverished populations and to avoid any kind of depression. However, following the caution principle, it seems important to have more reliable information on the behaviour and the ecology of the different populations to confirm definitively this lack of biologically significant separation.

Otherwise, eastern animals, safe from the Aleutian disease (e.g. the Romanian ones), could be used to reinforce the western population and released into the wild in France or in Spain. This cannot preclude a risk of outbreeding depression. Another way of proceeding should be to initiate a captive breeding programme with individuals from the western population and to release the offspring into areas formerly inhabited by the species but this does not offer any solution to the problem of the very low genetic variability.

Hence, we suggest to breed together western and eastern animals, namely those from the NE population because they are the more genetically diverse. The offspring should be reintroduced into regions of France formerly occupied by the western population and devoid of the American mink (*Mustela vison*). Should the reintroduced minks thrive, then the risk of outbreeding depression could be dismissed and it may be assumed that these 'mixed' minks could progressively reach the areas where western minks are still present. However, before any project of mink reintroduction in the wild, a more precise identification of the real causes at the base of the dramatic decline of the species seems of cardinal importance.

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Appendix

Geographic distribution and references of *Mustela* tissues used for the experiments

| Geographic origin                   | Total no. of animals | Sample symbols (see Figs 1, 2 and 3) | Tissue sample numbers or GenBank access (for sequenced samples) |
|-------------------------------------|----------------------|--------------------------------------|---|
| <i>Mustela lutreola</i>             |                      |                                      |   |
| France                              |                      |                                      |   |
| Aillas ou Braud et St louis         | 2                    | F1                                   | MLU 18 LA, MLU 09 GI  |
| Ambleville                          | 3                    | F2                                   | MLU16 gi, MLU 55 HP, MLU16hp                                    |
| Audenge                             | 2                    | F3                                   | MLU HP 39, MLU HP 52  |
| Bagas                               | 1                    | F4                                   | MLU 015 La  |
| Balizac                             | 2                    | F5                                   | MLU 007 GI, MLU 07 CHA  |
| Bazac                               | 1                    | F6                                   | MLU 018 GI  |
| Bégaar                              | 1                    | F7                                   | MLU 014 GI  |
| Belin Beliet                        | 2                    | F8                                   | MLU HP 56, MLU 003 GI   |
| Bellefond                           | 1                    | F9                                   | MLU 020 GI  |
| Bernos-Beaulac                      | 1                    | F10                                  | MLU 001 DO  |
| Blanquefort (Jalles de blanquefort) | 1                    | F11                                  | MLU 001 ORX   |
| Blaye                               | 1                    | F12                                  | MLU 002 D   |
| Bruges                              | 3                    | F13                                  | MLU 011 ORX, MLU P256, MLU 004 ORX                              |
| Bruges (Jalles de Blanquefort)      | 2                    | F14                                  | MLU 003 C, MLU 004 D  |
| Camiran                             | 1                    | F15                                  | MLU 001 D   |
| Crazannes                           | 1                    | F16                                  | MLU NERC 2  |
| Criteuil la Magdeleine              | 1                    | F17                                  | MLU 005 ORX   |
| Cubzac les Ponts                    | 1                    | F18                                  | MLU 012 ORX   |
| Cursan                              | 1                    | F19                                  | MLU 010 C   |
| Gastes                              | 1                    | F20                                  | MLU 005 C   |
| Guiche                              | 2                    | F21                                  | MLU 002 ORX, MLU 014 C  |
| Hontanx                             | 5                    | F22                                  | MLU 011 C, MLU 010 ORX, MLU 002 ENG, MLU 001 C, MLU 02 LG       |
| La Clisse                           | 1                    | F23                                  | MLU 001 Pa  |
| Labenne                             | 4                    | F24                                  | MLU 010 LA, MLU 009 LA, MLU 017 GI, MLU 017 LA                  |
| Lachaise                            | 3                    | F25                                  | MLU 002 GI, MLU 005 GI, MLU P238                                |
| Landeroute sur Ségur                | 1                    | F26                                  | MLU 009 CHA   |
| Léon                                | 4                    | F27                                  | MLU 007 LA, MLU 010 GI, MLU 004 LA, MLU P237                    |
| Les Billaux                         | 1                    | F28                                  | MLU P237  |
| Lugos                               | 1                    | F29                                  | MLU 011 GI  |
| Mezos                               | 1                    | F30                                  | MLU 024 LA  |
| Moliet et Maa                       | 3                    | F31                                  | MLU 022 LA, MLU 021 LA, MLU 004 NERC                            |
| Montpon-Ménésterol                  | 1                    | F32                                  | MLU 019 LA  |
| Morizes                             | 1                    | F33                                  | MLU 020 LA  |
| Nercillac                           | 2                    | F34                                  | MLU 023 LA, MLU 021 GI  |
| Neuffons                            | 1                    | F35                                  | MLU 016 LA  |
| Onard                               | 1                    | F36                                  | MLU 008 LA  |
| Orx                                 | 2                    | F37                                  | MLU 012 LA, MLU 003 E   |
| Parantis en Born                    | 1                    | F38                                  | MLU 005 D   |
| Parempuyre                          | 1                    | F39                                  | MLU 005 LA  |
| Perissac                            | 1                    | F40                                  | MLU 006 LA  |
| Pey                                 | 2                    | F41                                  | MLU 003 LA, MLU 012 GI  |
| Pont de Martrou                     | 2                    | F42                                  | MLU 008 GI, MLU 029 LA  |
| Pujo le Plan                        | 1                    | F43                                  | MLU 030 LA  |
| Puymiclan                           | 1                    | F44                                  | MLU 013 LA  |
| Roquebrune                          | 1                    | F45                                  | MLU 002 PA  |
| Saillans                            | 2                    | F46                                  | MLU P236, MLU 003 LG  |
| Sanguinet                           | 1                    | F47                                  | MLU 001 LA  |
| St André de Seignanx                | 1                    | F48                                  | MLU 002 DO  |
| St Etienne d'Orthes                 | 1                    | F49                                  | MLU 001 GE  |
| St Gein                             | 2                    | F50                                  | MLU 015GI, MLU 014 LA   |
| St Laurent des Hommes               | 1                    | F51                                  | MLU 002 LA  |
| St Laurent Médoc                    | 1                    | F52                                  | MLU002 C  |

## Appendix Continued

| Geographic origin          |                           | Total no. of animals | Sample symbols (see Figs 1, 2 and 3) | Tissue sample numbers or GenBank access (for sequenced samples)   |
|----------------------------|---------------------------|----------------------|--------------------------------------|---|
|                            | St Leger de Balson        | 3                    | F53                                  | MLU 004 GI, MLU 019 GI, MLU 008 CHA   |
|                            | St Louis de Montferrand   | 2                    | F54                                  | MLU 028 LA, MLU 025 LA  |
|                            | St Martial sur Né         | 2                    | F55                                  | MLU 001 GI, MLU 006 C   |
|                            | St Martin de Seignanx     | 1                    | F56                                  | MLU 3   |
|                            | St Médard d'Eyrans        | 1                    | F57                                  | MLU 02 05 00  |
|                            | St Médard en Jalles       | 1                    | F58                                  | MLU E   |
|                            | St Paul en Born           | 1                    | F59                                  | MLU VM8   |
|                            | St Vincent de Paul        | 1                    | F60                                  | MLU C   |
|                            | Ste Eulalie en Born       | 1                    | F61                                  | MLU 004 C   |
|                            | Ste Gemme                 | 1                    | F62                                  | MLU 060400  |
|                            | Tartas                    | 1                    | F63                                  | MLU VF5   |
|                            | Tizac de Lapouyade        | 1                    | F64                                  | MLU VM6   |
|                            | Tocanne St Apre           | 1                    | F65                                  | MLU VF7   |
|                            | Uzeste                    | 2                    | F66                                  | MLU D, MLU VF4  |
|                            | Vendays-Montalivet        | 1                    | F67                                  | MLU 001 Cm  |
|                            | Vensac                    | 1                    | F68                                  | MLU 003 CHA   |
|                            | Vielle st Girons          | 2                    | F69                                  | MLU 004 CHA, MLU 005 CHA  |
|                            | Villandraut               | 2                    | F70                                  | MLU 006 CHA, MLU 003 DROPT  |
|                            | Villefranque              | 1                    | F71                                  | MLU004 ORX  |
|                            | Villeteon                 | 1                    | F72                                  | MLU V1 (= mluv1montp)   |
| Spain                      | La Rioja                  | 2                    | Sp1                                  | Mulp 55, Mulp 56  |
|                            | Navarra                   | 1                    | Sp2                                  | Mulp 50   |
|                            | Gipuzkoa                  | 1                    | Sp3                                  | AF207725  |
|                            | Najerilla river; La Rioja | 1                    | Sp4                                  | Uruñuela  |
|                            | Ebro river; Álava         | 1                    | Sp5                                  | Tequila   |
|                            | Ayuda river; Burgos       | 1                    | Sp6                                  | Becaria   |
|                            | Aragon river; Navarra     | 1                    | Sp7                                  | ML14  |
|                            | Alhama river; Navarra     | 1                    | Sp8                                  | ML13  |
|                            | Ebro river; La Rioja      | 1                    | Sp9                                  | ML140   |
|                            | Ebro river; Burgos        | 1                    | Sp10                                 | ML141   |
|                            | Oca river; Burgos         | 1                    | Sp11                                 | Trini   |
|                            | Muxika locality; Bizkaia  | 1                    | Sp12                                 | Muxica1   |
|                            | Ebro river; La Rioja      | 1                    | Sp13                                 | Arrinconado   |
|                            | Bidasoa river; Navarra    | 1                    | Sp14                                 | Navarra3ML041E  |
| Romania                    | Danube delta              | 34                   | Ro1                                  | JRM-698, JRM-699, 2, 11, 20, 28, 42, 44, 45, 48, 51, 54, 56, 58, 59, 63, 64, 65, 68, 71, 73, 75, 77, 81, 82, 83, 90, 93, 94, 99, 102, 107, 111, 153 |
| Estonia                    | Tallin                    | 2                    | Es1                                  | JRM-659, JRM-660  |
|                            | Unknown locality          | 1                    | Es                                   | AF207723  |
| Bielorussia                | Vitebsk                   | 2                    | Bi1                                  | Mulu 3, AF207724  |
| Russia                     | Pskov                     | 2                    | Ru1                                  | Mulp 6, Mulp 7  |
|                            | Tver                      | 11                   | Rus2                                 | AF207720, Mulp 8 to Mulp 13, 811, 1011, 1111, 1025  |
| <i>Mustela putorius</i>    |                           |                      |                                      |   |
| Spain                      | La Rioja                  | 1                    | Sp1                                  | AF207726  |
|                            | Gipuzkoa                  | 1                    | Sp3                                  | Mulp 53   |
| Portugal                   | Unknown locality          | 1                    |                                      | MPU 27  |
| Estonia                    | Unknown locality          | 2                    |                                      | Mulp 2, Mulp 3  |
| Russia                     | Unknown locality          | 1                    |                                      | Mulp 41   |
|                            | Tver                      | 1                    | Rus2                                 | Mulp 35   |
| ?                          |                           | 1                    |                                      | AF207717  |
| ?                          |                           | 1                    |                                      | AF207718  |
| ?                          |                           | 1                    |                                      | AF068570  |
| <i>Mustela eversmannii</i> |                           |                      |                                      |   |
| Mongolia                   | E. Inner                  | 1                    |                                      | M. Evers. 1   |
| Serbia                     | Northwest region          | 1                    |                                      | M. Evers. 2   |
| <i>Mustela nigripes</i>    |                           |                      |                                      |   |
| USA                        | Unknown locality          | 2                    |                                      | M. nigripes 1 and 2   |